

# IMPEDANCE SPECTROMETRY FOR MONITORING ALCOHOLIC FERMENTATION KINETICS UNDER WINE-MAKING INDUSTRIAL CONDITIONS

*Miguel A. Pérez*<sup>1</sup>, *Rocío Muñiz*<sup>1</sup>, *Cristina de la Torre*<sup>1</sup>, *Beatriz García*<sup>1</sup>, *Carlos E. Carleos*<sup>2</sup>, *Raúl Crespo*<sup>3</sup>, *Luis M. Cárcel*<sup>3</sup>

<sup>1</sup>Dept. de Ingeniería Eléctrica (University of Oviedo), Gijón, Spain, [maperezg@uniovi.es](mailto:maperezg@uniovi.es)

<sup>2</sup>Dept. de Estadística e Investigación Operativa (University of Oviedo), Gijón, Spain, [carleos@uniovi.es](mailto:carleos@uniovi.es)

<sup>3</sup>Dept. de Ingeniería Agroforestal (University of Valladolid), Palencia, Spain, [lmcarcel@iaf.uva.es](mailto:lmcarcel@iaf.uva.es)

**Abstract** – This paper presents a system for monitoring several parameters during alcoholic fermentation in wine industry. Alcoholic fermentation of grape juice is a quite complex biological process that involves a large number of variables and boundary conditions. The number of cells and total production of carbon dioxide are two important parameters for monitoring this process and several decisions must be carried out depending on these values. This paper presents an on-line application of impedance spectrometry to estimate both, CO<sub>2</sub> and cells in a small-scale fermentation plant. Experimental results has been obtained from several micro-fermentation, verifying the capability of carbon dioxide measurement and the estimation of yeast cell concentration.

**Keywords:** impedance spectrometry, fermentation, wine

## 1. INTRODUCTION

Wine making from grape juice to finished wine is a complex process that involves several stages, often conducted empirically and traditionally. Most important steps are the skin maceration and the alcoholic fermentation that determines final organoleptic characters. Alcohol production during fermentation process can be determined by measurement of total carbon dioxide release [1], [4]. So, fermentation kinetics could be controlled (or followed) by measurement of total carbon dioxide.

Carbon dioxide and alcohol production is a consequence of biological activity of cells during fermentation stage. Thus, to improve control actions, total number of cells should be obtained in addition to carbon dioxide production. Variables, CO<sub>2</sub> and total cell count should be on-line measured in order to provide criteria for process controlling.

On-line measurement of carbon dioxide in liquid (grape juice to wine) can be carried out by means of resistance, due to effect of CO<sub>2</sub> bubbles (carbon dioxide bubbles increase the value of total resistance). Cell count affects to real permittivity [2], [3] and can be obtained by capacitance measurement at appropriate frequency.

This paper describes a system for monitoring both, cell count and CO<sub>2</sub>, by using impedance spectrometry; in next sections, a brief system description and several experimental results will be presented.

## 2. MEASUREMENT SYSTEM

Measurement system has been included into a full-controlled micro-fermentation plant as it is shown in Fig. 1 able to deliver 1000 to 2000 ml of grape juice. It consists in a vessel that contains the grape juice and an impedance cell for liquids from Hewlett-Packard (HP16452A). Liquid circulates from bottom of vessel to impedance cell and from this one to the vessel by means a controlled peristaltic pump to avoid any change in bubbles production.

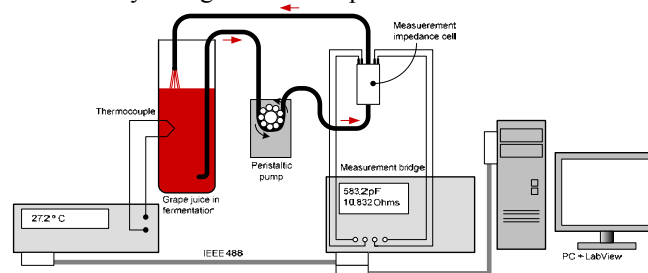


Fig. 1. Micro-fermentation plant including on-line measurement of carbon dioxide and cell count by means of impedance spectrometry.

Capacity and resistance values are calculated in a measurement bridge (impedance analyzer from Hewlett-Packard (HP4284A) and liquid temperature is also measurement by means of a K-type thermocouple to compensate its effect on resistance measurement. All measured values are transmitted via IEEE-488 to a standard PC that includes a LabView program able to store, display and manage all data.

Fig. 2 shows the impedance measurement cell and its electrical equivalent circuit when grape juice is the liquid inside: a parallel circuit with a capacitance,  $C_p$  and a resistance,  $R_p$ . But, grape juice has a lot of dissolved ions

and consequently, it has a high value of conductivity. So,  $R_p$  produces a low value – around 10 Ohms for the particular geometry of impedance cell, characterized by a distance between electrodes of 2.6 mm – introducing additional problems in capacitance estimation because  $R_p$  seems a short-circuit. An increase of electrodes distance of measurement cell will increase the value of  $R_p$  but it would reduce of measured capacity  $C_p$ .

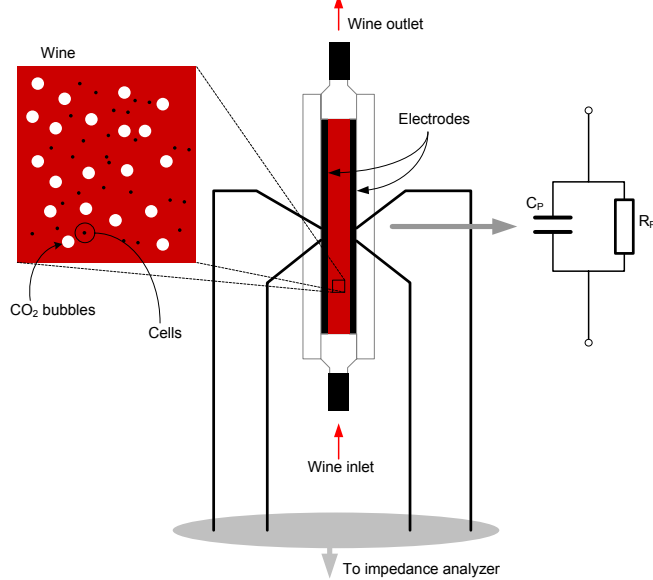


Fig. 2. Impedance measurement cell with grape juice (or wine) inside connected to impedance analyzer modelled with a resistive-capacitive equivalent circuit. Carbon dioxide and cells in liquid affects to equivalent circuit parameters.

The effect of carbon dioxide can be well modelled by liquid conductivity, that is, carbon dioxide bubbles reduce the effective section of liquid conductor in impedance cell, increasing total resistance. Additional effects, such as changes in intrinsic conductivity due to liquid composition modification become negligible in relation to large effect of bubbles. Conductivity in liquid is measurement with AC excitation to avoid polarization effects caused by dissolved ions,

The effect of any kind of cell over real permittivity of liquid and, consequently, over capacity value, has been well established for other high-conductivity liquid such as raw milk with presence of somatic cells [2], [3]; the quantitative effect over capacity is depending on cell size and excitation frequency.

### 2.1. Impedance spectrometry during wine-making

The micro-fermentation workbench in Fig. 1 can be used for any kind of impedance spectrometry of liquids, but must be adapted to the particular conditions of wine-making.

Under the special conditions of grape juice fermentation, and when *Saccharomyces Cerevisiae* is used as yeast, excitation frequency must range from 300 kHz to 1 MHz for capacity measurement and around of 40 kHz for resistance (or conductivity) measurement. However, no significant differences were found in resistance measurement when excitation frequency ranges from 4 to 400 kHz.

### 2.2. Thermal effects

Temperature is a controlled variable in industrial wine-making process because it affects to the fermentation kinetics and other processes. In our laboratory-level plant for micro-fermentation, temperature was not controlled and its evolution depended on room conditions; it needs a minimum value to start and to hold the fermentation and this minimum was guaranteed.

However, monitoring system of Fig. 1 reads the temperature of grape juice, because the conductivity – and electrical resistance – of liquids has a heavy thermal-dependence. There are models for thermal dependence of conductivity of pure water and ultra-pure water (high resistance liquids) but these models are no valid for high-conductivity liquids such as wine and grape-juice. For this case we will use a linear model because temperature range is enough short to consider non-linearity effect as negligible.

$$R_T = R_{TREF}(1 + \alpha(T - T_{REF})) \quad (1)$$

where  $R_T$  is the measured resistance at the measured temperature  $T$ ,  $R_{TREF}$  is the resistance at  $T_{REF}$ , and  $\alpha$  is the thermal coefficient.

## 3. EXPERIMENTAL RESULTS

Several micro-fermentation processes have been carried out by using the system shown in Fig. 1 with 1500 ml of “tempranillo” grape juice with 0.8 g of *Saccharomyces Cerevisiae* and 1 g of fermentation activator. In all fermentation processes, total time involved was 10 days (248 hours), time enough to complete fermentation [5], [6] under the specific conditions of this experiment. The evolution of variables is stored by means a custom LabView program with 30 min of sample period. In all cases, sampled variables are: liquid temperature, electrical capacity,  $C_p$  and parallel resistance,  $R_p$  of equivalent electrical model shown in Fig. 2 for liquid impedance cell. These values are measured from 100 Hz to 1 MHz with a logarithmic sweep.

All measured variables exhibit similar behaviour in all fermentation processes carried out; for example, Fig. 3 shows the value of measured resistance,  $R_p$  at 40 kHz of excitation frequency– found values of  $R_p$  are quite similar from 400 Hz to 400 kHz – and Fig. 4, the liquid temperature during a complete fermentation.

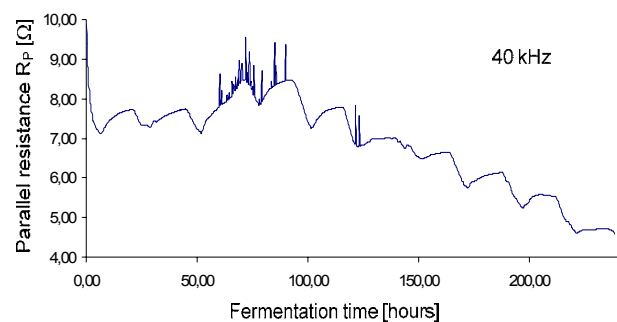


Fig. 3. Evolution of liquid resistance,  $R_p$  at 40 kHz during fermentation stage of wine-making (240 hours).

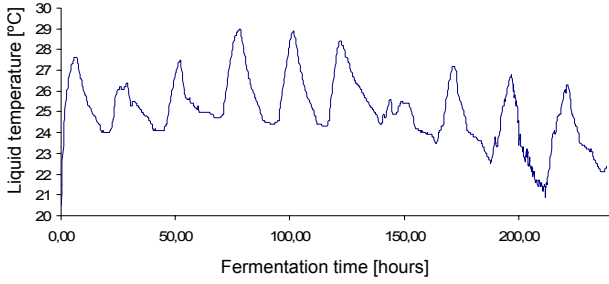


Fig. 4. Liquid temperature during fermentation stage (240 hours).

Thermal effects on measured resistance of high-conductivity liquids can be compensated by means an expression such as (1). In the case of grape juice, thermal coefficient  $\alpha$  in that equation can be calculated from the micro-fermentation workbench of Fig. 1, using values where changes are due to temperature because liquid has not biological evolution: Fig. 5 shows the relationships between temperature and resistance in that case and we can verified a high correlation when the thermal model of (1) is used.

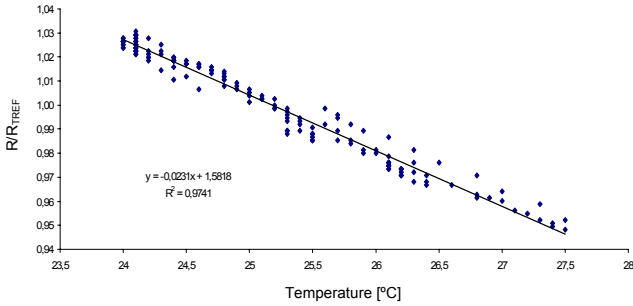


Fig.5. Relationship between temperature of liquid a normalized resistance  $R/R_{TREF}$ . A high correlation coefficient was found for a model similar to (1).

The obtained thermal model from Fig.5 can be presented as:

$$R_T = R_{25^\circ C} (1 - 0.0231(T - 25^\circ C)) \quad (2)$$

where  $R_T$  is the measured resistance at temperature  $T$ . Similar empirical procedure applied to other types of grape juice has produced similar values of the thermal coefficient  $\alpha$ , so, it could be considered as a constant for grape juice.

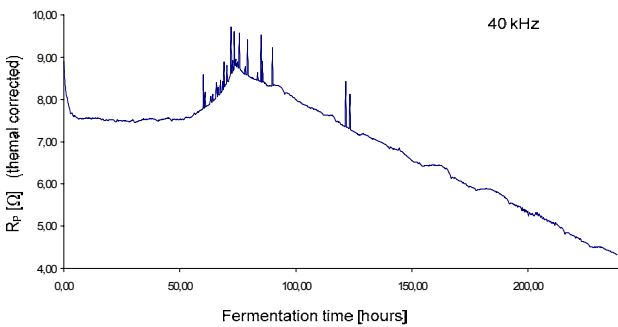


Fig. 6. Evolution of liquid resistance,  $R_P$  at 40 kHz (thermal correction is applied) during fermentation stage of wine-making (240 hours).

Values of measured parallel resistance of Fig. 3 has been corrected by means of expression (2), producing the curve of Fig. 6.

Parallel resistance value,  $R_P$  after temperature correction exhibits similar behaviour to carbon dioxide production in alcoholic fermentation process [5], [6], presenting a maximum value when the fermentation produce the maximum carbon dioxide release. This effect is also verified by notable visual effects such as a huge production of bubbles in liquid and tumultuous fermentation in grape vessel.

The value of capacity  $C_p$  during fermentation presents dependence on frequency as it can be seen in Fig. 7 but all of cases exhibits similar evolution including some influence of liquid temperature.

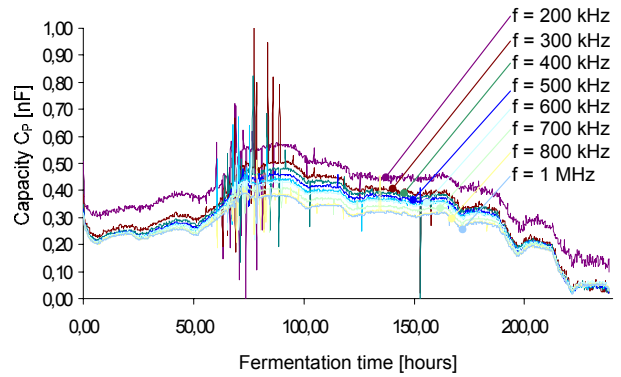


Fig. 7. Evolution of capacity,  $C_p$  during fermentation stage of wine-making (240 hours) in function of excitation frequency.

Speaks in measured value are due to perturbation caused by presence of huge  $CO_2$  bubbles, and could be removed by a simple averaging.

The evolution of measured capacity,  $C_p$  – at any frequency from 200 kHz to 1 MHz – is similar to the evolution of yeast cell concentration [5], [6]: after 50 hours of beginning, the number of yeast cells increase up to its maximum concentration during tumultuous fermentation and then, it decreases slowly to the end of fermentation. The evolution of measured resistance,  $R_P$  at any excitation frequency completes the information about fermentation kinetics but, in this case, is focused in carbon dioxide release. Both values, capacity  $C_p$  and resistance  $R_P$  are plotted in Fig. 8 to show how they can explain the fermentation kinetics:

- First stage: from process beginning, the total count of yeast cells increase slowly and there is not visible carbon dioxide bubbles. This stage is characterized by a constant value of liquid resistance and a slow increase of capacity.
- Fermentation starts with a multiplication of yeast cells and they begin to transform sugar in alcohol, causing an increase of  $CO_2$  release. This instant is defined by a hard increase in slope of  $R_P$  and  $C_p$ .
- Tumultuous stage: there are a lot of yeast cells and they are extremely active, causing the maximum release in carbon dioxide.  $R_P$  reaches it maximum value and  $C_p$  keeps in high values.

- After the tumultuous stage, fermentation continues, but the CO<sub>2</sub> release decrease due to the consumption of sugar and the cell count begins to fall. These effects are represented, respectively by the decrease of C<sub>p</sub> and R<sub>p</sub>.
- End of fermentation, without gas release because most of sugar has been yet transformed in alcohol. C<sub>p</sub> and R<sub>p</sub> have their minimum values.

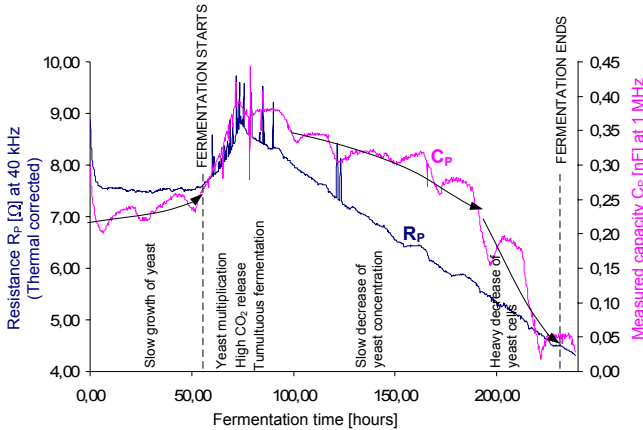


Fig. 8. Evolution of capacity, C<sub>p</sub> during fermentation stage of wine-making (240 hours) in function of excitation frequency. Speaks in measured value are due to perturbation caused by presence of huge CO<sub>2</sub> bubbles, and could be removed by a simple averaging.

#### 4. A NEW CARBON DIOXIDE SENSOR BASED UPON IMPEDANCE SENSOR

A new carbon dioxide sensor has been designed according to the results obtained from workbench of Fig.1 during micro-fermentation. This sensor consists of two plates of stainless steel 304 sunken in liquid, those impedance is measured by means an AC Wheatstone bridge excited at 40 kHz with thermal correction following (2). The equivalent model is not a pure-resistive circuit and some capacitive effect must appear, but the distance between both plates (electrodes) is enough to reduce that disturbance. Thus, impedance measured will be similar to liquid resistance between two plates.

This sensor and the conditioning circuit have been used under industrial condition fermentation and their results are compared to the values produced by a commercial CO<sub>2</sub> sensor. Fig. 9 shows this comparison.

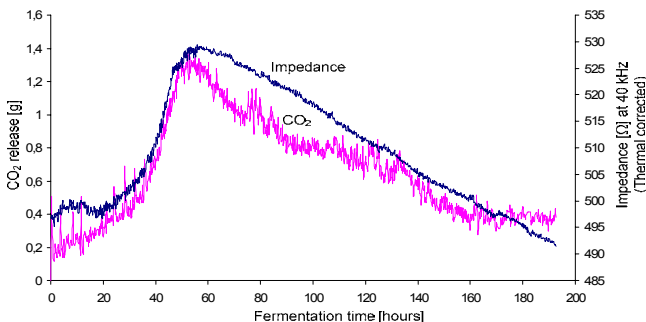


Fig. 9. Comparison of carbon dioxide release and measured impedance in an industrial fermentation.

As we can see, the values of impedance sensor exhibits a similar evolution that values provided by a commercial CO<sub>2</sub> sensor, with perfect agreement in determination of fermentation start time and the instant of maximum gas release. Small differences were found after maximum gas release point, due to the additional capacitive effect of yeast cells.

Fig. 10 plots the value of gas release in function on measured impedance and we can found a high correlation that can validate this kind of sensor.

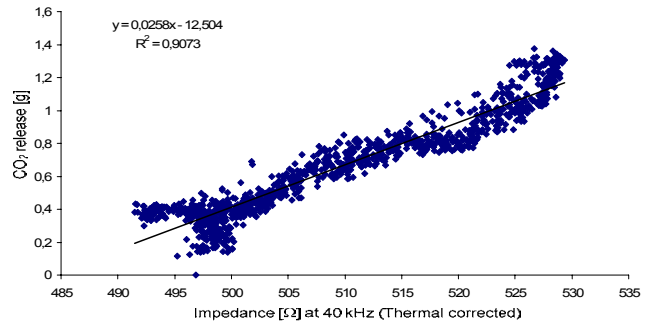


Fig. 10. Use of liquid impedance to model CO<sub>2</sub> release.

But, if we verify the time evolution of two variables – carbon dioxide release and resistance – we can see that both parameters have a similar behaviour except for a small area after the maximum value of gas release, when the concentration of yeast cells reaches its maximum value, as we can see in Fig. 10.

The interest of the estimation of CO<sub>2</sub> is due to the relationship between the evolution of must mass that controls the moment of fermentation and the carbon dioxide release as we can see in the Fig. 11.

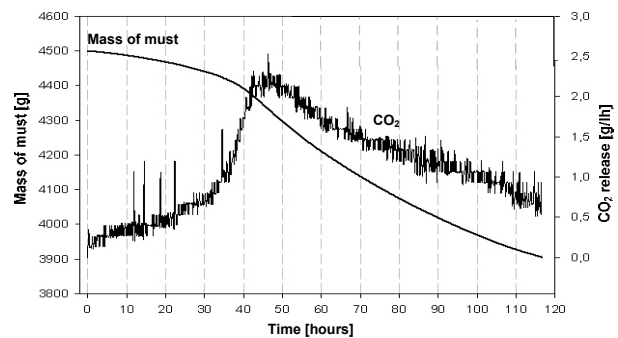


Fig. 11. Comparison of carbon dioxide release and the must quantity to demonstrate why the gas release is a very interesting variable to predict the kinetic of fermentation process.

#### 5. CONCLUSIONS

Analysis of resistance and capacitance can be used to estimate the kinetics of alcoholic fermentation during wine-making process. After the results obtained from a micro-fermentation plant, measured resistance after temperature compensation can be used to estimate carbon dioxide release, and the total number of yeast cells could be estimated with capacitance at high frequencies (0.2 to 1 MHz). Finally, a CO<sub>2</sub> sensor based upon liquid impedance

has been designed as a consequence of experimental results. This sensor has been verified during real fermentation process under industrial conditions and provide similar results that very expensive commercial CO<sub>2</sub> sensor.

#### ACKNOWLEDGMENT

This work has been supported by R&D project Ref. DPI2005-09200-C02 of Spanish Gov.

#### REFERENCES

- [1] N.E. El Haloui et al. Method for on-line prediction of kinetic of alcoholic fermentation in wine making, *J. of Fermentation and Bioengineering* Vol. 68, No 2, pp. 131-135, 1989.
- [2] G.J. Grillo et al. Design of a Low Cost Mastitis Detector in Cows by Measuring Electrical Conductivity of Milk. *IEEE Instr. and Meas. Tech. Conf.* Volume 1, pp. 375 - 378. 2002.
- [3] G.J. Grillo et al. *Method and System for Evaluation of Sub-Clinical Mastitis Level in Fresh Cow Milk*, ES patent no. 20050916, 2005.
- [4] R. Táboas et al. Study of the alcoholic fermentation in wine Albariño. Influence of yeast *Saccaromyces* and SO<sub>2</sub>, *J. Environmental, Agricultural and Food Chemistry* 1 (2), 126-136. 2002.
- [5] S. Malherbe et al. Modelling effects of assimilable nitrogen and temperature on fermentation kinetics on enological conditions", *Biotechnology and Bioengineering* Vol.86, 3. 261-272, 2004.
- [6] J.M. Salmon, Enological fermentation kinetics of a isogenic ploidy series derived from an industrial *Saccharomyces Cerevisiae* strain, *J. of Fermentation and Bioengineering* Vol. 83, No 3, 253-260. 1997.